

Expert Opinion

1. Introduction
2. Low-density lipoprotein-type carriers
3. High-density lipoprotein as a drug delivery vehicle
4. Enhanced targeting of lipoprotein–drug complexes via covalent modification
5. Comparison of lipoprotein-mediated drug delivery with other formulations
6. Conclusions
7. Expert opinion

informa
healthcare

Prospects and challenges of the development of lipoprotein-based formulations for anti-cancer drugs

Andras G Lacko[†], Maya Nair, Laszlo Prokai & Walter J McConathy

[†]University of North Texas Health Science Center, Department of Molecular Biology and Immunology, 3500 Camp Bowie Blvd, Fort Worth, TX 76107, USA

This review evaluates drug delivery systems that involve intact plasma lipoproteins or some of their components. These complex macromolecules transport highly water-insoluble compounds (cholesteryl esters and triacylglycerols) in their natural environment – a property that renders them ideal carriers of hydrophobic drugs. Particular emphasis is placed on the application of lipoproteins as drug delivery agents in cancer chemotherapy. The history and present activity regarding lipoprotein-based formulations are reviewed, with the primary focus on the smaller sized (low and high density) lipoprotein-based formulations and their potential clinical and commercial value. The use of both native and synthetic lipoproteins as drug delivery agents are discussed from the standpoint of therapeutic efficacy, as well as commercial feasibility. The advantages of lipoprotein-based drug delivery formulations are compared with other drug delivery models, with the primary focus on liposomal preparations. Finally, an expert opinion is provided, regarding the potential use of lipoprotein-based formulations in cancer treatment, taking into consideration the major advantages (biocompatibility, safety, drug solubility) and the barriers (manufacturing protein components, financial interest, investments) to their commercial development.

Keywords: cancer chemotherapy, drug delivery, lipoprotein, targeting

Expert Opin. Drug Deliv. (2007) 4(6):665–675

1. Introduction

Many intravenously administered pharmaceutical preparations, particularly anti-cancer drugs, perform with less than optimal efficiency because of limited accessibility to target tissues, systemic toxicities and tumor drug resistance [1]. An additional concern about developing new cancer chemotherapeutic agents is the inability of specific drugs to fully penetrate individual tumors [2]. The expectations for adjuvant therapy, despite recent improvements, are thus only modest, as many recurring cancers tend to be fatal because the presently available approaches are unable to eradicate the metastatic lesions [2]. Targeted anti-cancer agents – an advanced version of systemic therapeutic strategies – are considered a promising new frontier for treating cancer, offering perhaps the best hope for victory over this lethal disease [3–5]. Plasma lipoprotein-based nanoparticle carriers, in the present author's view, offer one of the best opportunities for designing and developing effective strategies for treating and eventually curing cancer. Consequently, this review deals with the relative merits of lipoproteins as drug carriers and the challenges involved in their developments as delivery vehicles for anti-cancer agents.

1.1 Lipoprotein carriers as vehicles for targeted cancer chemotherapy

The structural characteristics and metabolic functions of lipoproteins have been extensively reviewed [6-8] and, thus, will be referred to only briefly here. Plasma lipoproteins are spherical macromolecular complexes of specific apolipoprotein and lipid components (Figure 1A) that transport water-insoluble lipids to their specific organ/tissue destinations.

The basic structure of lipoproteins involves an outer protein-phospholipid shell with a lipophilic surface and an interior hydrophobic compartment (Figure 1A) for the transport of water-insoluble lipid components (triacylglycerols and cholesteryl esters). This structural arrangement is also eminently favorable for the transport of hydrophobic drugs (e.g., paclitaxel) that are often used in cancer chemotherapy (Figure 1B). In the context of being able to transport hydrophobic molecules, lipoproteins are, thus, attractive candidates for delivering anti-cancer agents to tumors, due to their bio-compatible components and the opportunity for targeted delivery by receptor-mediated uptake of the drug-loaded low-density lipoproteins (LDLs) by endocytosis [2,10] or by the selective uptake of high-density lipoprotein (HDL) core components [11,12]. Both types of these lipoprotein receptors are overexpressed by malignant cells and tumors [9,12,13-17]. Therefore, lipoproteins are expected to increase the therapeutic efficacy by targeting drugs to tumors that overexpress lipoprotein receptors and, thus, minimize systemic toxicity by shielding the drug from contact with most normal tissues [18]. In this review, the authors limit the discussion to two classes of lipoprotein carriers based on the structure of LDLs and HDLs.

2. Low-density lipoprotein-type carriers

2.1 Circulating (native) low-density lipoprotein

Initially, Krieger *et al.* [10,19] replaced the core of native LDL with cholesteryl linoleate and concluded that a broad range of hydrophobic compounds, including drugs, may thus be incorporated into LDL [10]. Subsequently, Gal *et al.* [15] proposed the use of LDL as a delivery vehicle for radiological or chemotherapeutic agents to gynecological neoplasms. More recently, Masquelier *et al.* have attempted to use LDL [20] and similar emulsions [21] as a vehicle for delivering chemotherapeutic agents to leukemic cells. Although these studies are promising, having shown solubilization of hydrophobic drugs and the overcoming of drug resistance, to a degree, in leukemic cells [22], the lack of *in vivo* efficacy data suggest that these approaches are still relatively early in their development. Kader and Pater [23] prepared drug-loaded LDL, as well as HDL, and concluded that incorporation of the drug did not change the structure of the lipoprotein particle, and the cytotoxicity of the encapsulated drug increased substantially.

De Smidt and Van Berkel reviewed the methods of preparation and properties of a number of LDL-based drug

formulations and concluded that these preparations represent 'an advanced biotechnological system whereby natural endogenous pathways are utilized for site-specific drug delivery' [24]. Nevertheless, concerns remained regarding the appropriate selection of the drug to be incorporated into LDL, and methods for the encapsulation of hydrophobic drugs into the core of the LDL complex. The design of LDL-drug complexes for the purpose of drug delivery to cells, particularly to cancer cells, has been based on the premise that rapidly proliferating cells and tissues have an enhanced need for cholesterol and, thus, have a higher expression of LDL receptors for increased LDL internalization [9,16,25]. Although this is a rational hypothesis, it has not been proven to be universally correct. Accordingly, although most malignant cells show higher LDL receptor expression than the corresponding normal cells [9,16,25-27] some show decreased expression or abnormal behavior [28-30]. In addition, receptor expression did not always correlate with, internalization of LDL – a key feature of the mechanism for projected applications in chemotherapy [31].

In summary, although cancer cells generally exhibit an increase in LDL receptor function, the overexpression of LDL receptor protein in tumor cells has not been consistently documented.

Vitols *et al.* conducted the most extensive studies, utilizing LDL particles as potential drug carriers in leukemia chemotherapy [25,27,32-36]. Their approach was based on findings that leukemia patients had substantially reduced LDL levels and enhanced LDL receptor activity in leukemia cells [33,35]. These studies led to the incorporation of the lipophilic drugs AD-32 [32] and WB4291 [34] into LDL and the assessment of their respective bio-distributions [32] and anti-leukemia activity [34,36]. Subsequently, several anthracycline derivatives were studied [37], resulting in LDL-drug complexes with some, but a number of these hydrophobic compounds proved to be difficult to incorporate into LDL. The overall stability of the drug-carrying LDL particle, upon injection into the plasma, proved to be less than satisfactory, with these formulations precluding more extensive *in vivo* and clinical studies [36,38]. Another approach by Lo *et al.* and Fung *et al.* produced LDL-doxorubicin complexes that displayed reduced toxicity to normal tissues, but were nearly as toxic to malignant tumors as the free drug [39], and exhibited reduced drug resistance in R-HepG2 cells [40].

LDL has been used for solubilizing and targeting a number of additional compounds of pharmaceutical interest, including photosensitizers [41,42], nucleosides [43], isoflavones [44], acrylophenone [45], anti-HIV drugs [46] and fluorescent imaging agents [47]. Nevertheless, the clinical application of native LDL particles as drug delivery agents is complicated by the procedures required for their isolation [24], methods of incorporation of the therapeutic agent [20], and related safety concerns [24,48]. The difficulties encountered in the use of native LDL-drug complexes for therapeutic purposes include the adequate supply of LDL,

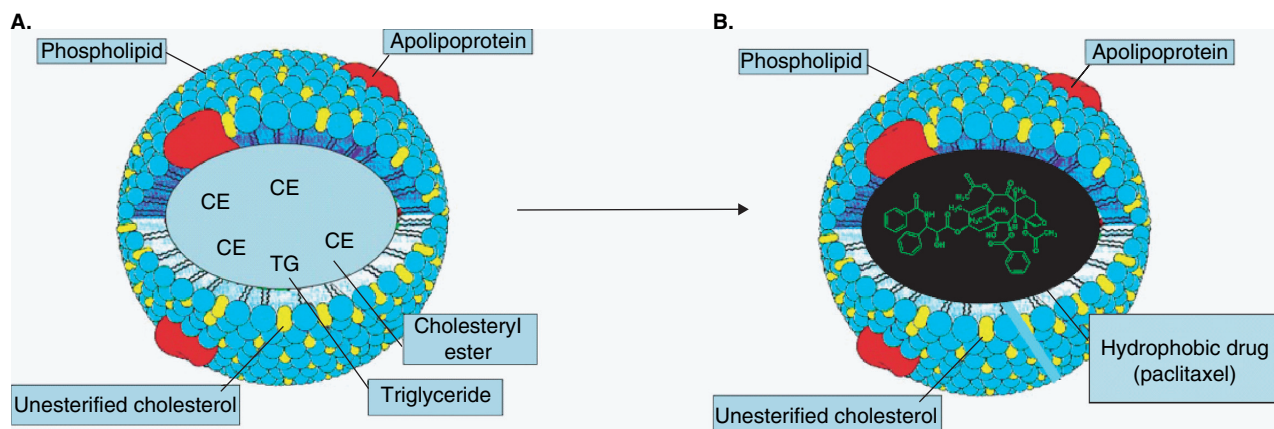


Figure 1. Localization of a hydrophobic drug (paclitaxel) in the core region of a generic lipoprotein.

Reproduced from WEINBERG RB: Lipoprotein metabolism: hormonal regulation. *Hosp. Pract. (Off Ed.)* (1987) **22**(6):223-227, 230, 233-234.

and its tendency to aggregate, as have been summarized by Owens *et al.* [49] and Shaw and Shaw [50]. These issues, despite the compelling biological rationale [19,24], render the use of native LDL impractical as a drug delivery agent in a clinical setting. Consequently, alternate strategies have been developed via artificially prepared, reconstituted LDL for the purpose of targeted drug delivery.

2.2 Synthetic (reconstituted) low-density lipoprotein, including apolipoprotein E-containing particles

The major challenge to the design and preparation of reconstituted low-density lipoprotein (rLDL) is the isolation of a soluble lipid-free apolipoprotein B-100 – the major protein component of the circulating LDL complex [7]. Some of the difficulties in involving apolipoprotein B-100 in synthetic formulations are associated with its high molecular weight and its tendency to aggregate upon delipidization. The early attempts to prepare rLDL, the methods for its production and the physical/chemical properties of the resultant particles have been reviewed [51,52]. A number of investigators attempted to prepare synthetic LDL/rLDL as a drug carrier, despite the limitations posed by the availability of apolipoprotein B-100 [7]. Lundberg [16,53,54] combined egg yolk phosphatidylcholine, a hydrophobic drug and apolipoprotein B-100 [15], resulting in a 23-nm rLDL particle that was taken up by the LDL receptor of cultured fibroblasts [16].

Van Berkel and coworkers prepared unilamellar liposomes, containing apolipoprotein E that were taken up by rat tissue LDL receptors [55], and similar particles containing a lipophilic derivative of daunorubicin that were taken up by B16 tumors [56]. By combining egg yolk phosphatidylcholine, triolein, cholesterol and cholesteryl oleate, Maranhao *et al.* developed a protein-free microemulsion (LDE) that was shown to be able to carry lipophilic drugs [57]. These LDE particles acquired apolipoprotein E upon injection into rats and were cleared at a rate similar to LDL. When the LDE particles were loaded with oleoyl-etoposide [58] and

oleoyl-paclitaxel [59], studies in mice showed an improvement in the therapeutic efficacy of encapsulated oleoyl-paclitaxel compared with the free drug. The metabolism of the LDE particles was studied in gynecological cancer patients, where the tumor tissue incorporated 3.5-times more paclitaxel-oleate from the LDE particles than normal tissues [60].

To circumvent the need for intact apolipoprotein B in the preparation rLDL–drug complexes, Baillie *et al.* utilized peptides covering the receptor binding region of apolipoprotein B to produce drug-carrying nanoparticles that supported the growth of U937 human lymphoma cells similar to native LDL [61]. More recently, Nikanjam *et al.* [62] prepared a synthetic LDL using peptides representing the lipid binding motif and the LDL-receptor binding domain of apolipoprotein B-100. The fluorescently labeled “nano-LDL” particles [33] were taken up by glioblastoma cells by the LDL receptor-mediated mechanism. The findings of these studies support the concept that targeted delivery of anti-cancer drugs by lipoprotein-type carriers via upregulated receptors in cancer cells and tumors [17,27] is feasible and, thus, could pave the way for clinical studies using these nanoparticles.

3. High-density lipoprotein as a drug delivery vehicle

During the initial development of lipoprotein-based drug carrier systems, the primary focus was centered on LDL because of the elegant, detailed characterization of LDL receptor structure and function [63], and the finding that rapidly proliferating cells (cancer cells) had enhanced LDL receptor expression [16,24], resulting in mild hypocholesterolemia in cancer patients [64]. However, recent studies, based on data from 530 cancer patients, revealed that HDL cholesterol levels were impacted to approximately the same extent (-13% for LDL-cholesterol, -12% for HDL-cholesterol) by malignancies [65]. Similar findings were reported for patients with hematologic cancers [66-68],

small cell lung cancer [69] and colorectal adenoma [70]. The characterization of the receptors (scavenger receptor B1 and CD36, and LIMPII analogous-1 [CLA-1]) that facilitate the selective uptake of cholesterol esters from HDL [71-73] have provided further impetus for the development of HDL-based, targeted drug delivery vehicles [18].

A number of earlier studies indicated that HDL may be an attractive drug carrier, particularly in cancer chemotherapy as the proliferation of adenocarcinoma [74] and other malignant cells have been shown to be accelerated by HDL or HDL components [75,76]. More recently, Pussinen *et al.* have shown that breast cancer cells acquire cholesterol from HDL via the CLA-1 receptor pathway [11], and Cao *et al.* reported that a mutant form of CLA-1 receptor suppressed the growth in a breast cancer cell line [17]. These data, combined with other reports, strongly suggest that the HDL receptors may play a role in the proliferative capacity of cancer cells and tumors that is at least as important as that of the LDL receptor [9,10,27]. Furthermore, the recent focus on HDL receptors in the biology of cancer cells [77,78] provides strong impetus for the development of HDL or HDL-type complexes as anti-cancer drug delivery agents. The concept of the selective targeting of HDL or reconstituted HDL (rHDL) drug complexes to malignant cells and tissues has been supported by data from Cao *et al.*, who showed that, in mice carrying MCF7 (breast) tumors, the tumor cells were expressing markedly higher levels of the CLA-1 receptor compared with the surrounding normal cells [17]. These findings are consistent with earlier data from the present authors' laboratory showing that cancer cells produced substantially stronger immunoblots with an antibody for the scavenger receptor B1 than normal fibroblasts [12]. It has also been shown that the uptake of paclitaxel and cholesteryl esters from rHDL particles was highly correlated ($r^2 = 0.88$; $p = 0.04$), suggesting that paclitaxel was taken up by cancer cells via the HDL receptor mechanism from the rHDL-paclitaxel complex [12].

3.1 Native (circulating) high-density lipoprotein

The utilization of native HDL as a drug delivery vehicle is fraught with the same difficulties as listed for LDL above, namely the difficulty of large scale isolation, bio-safety issues and consequently the excessive cost involved in the development of the HDL-drug formulations [24]. As a result, only a limited number of attempts have been made to incorporate drugs into native HDL [23], and none with the ultimate purpose of therapeutic applications. The following section reviews the application of a more appropriate model, rHDL, for an effective drug delivery system.

3.2 Synthetic (reconstituted) high-density lipoprotein

One of the confounding issues regarding rHDL relates to the exact definition of what is meant by 'rHDL'. The terms 'reconstituted' and 'recombinant' have both been used in

the literature, although the former is more predominant recently, as 'recombinant' is now preferred for designating DNA constructs.

The nomenclature is quite clear for all other classes of lipoproteins; however, for rHDL, it is complicated by the nature of HDL metabolism, which has been reviewed in detail recently [79,80]. Specifically, discoidal precursor particles, composed primarily of phosphatidylcholine and apolipoprotein A-I – the major components of HDL – are secreted by the liver [81] and the intestine, and are subsequently rapidly converted to spherical particles [82] via the action of the enzyme lecithin:cholesterol acyltransferase in the blood (Figure 2).

Scientific publications [83,84], as well as patent disclosures [201] have referred to both discoidal and spherical particles occasionally as rHDL [83,84], but while some prefer to specify the nature of the lipoprotein complex by inserting discoidal or spherical designation before rHDL [85,86]. This unsystematic, alternative naming of rHDL particles can result in considerable confusion and misunderstandings, during the review of scientific articles, grants and patent proposals. The reader is, thus, alerted to the alternate naming of rHDL particles used by many of the articles referred to in this review.

For the purpose of formulating drug delivery complexes, spherical-shaped particles have a clear advantage over other configurations, as these have a defined core region to accommodate the drug to be transported (Figure 1). Although other interactions are possible between the hydrophobic drug and specific lipid components within the lipoprotein structure, the localization of the lipophilic drugs in the core region is preferred, due to enhanced stability of the rHDL-drug complex. Van Berkel and coworkers were the first to report on comprehensive studies of spherical 'neo-high density lipoproteins' that were proposed for use as drug delivery vehicles [87,88]. They incorporated the lipophilic prodrug iododeoxyuridine into neo-HDL and found that the molecular characteristics of the neo-HDL-drug complex were similar to that of circulating apolipoprotein E-free HDL [88]. Studies from the present authors' laboratory have shown that a stable rHDL-drug complex could be prepared by combining Taxol® (Bristol-Myers Squibb) and the components from circulating HDL [12] via a procedure similar to that described by Van Berkel and coworkers [88]. The rHDL-Taxol complex was taken up by cancer cells, apparently by a receptor-mediated mechanism, resulting in the killing of the cancer cells by the incorporated drug [12]. A more advanced rHDL formulation, containing paclitaxel, has been developed recently that has substantially increased drug-carrying capacity and cytotoxicity toward several cancer cells [91]. Recent studies by Lou *et al.* resulted in the preparation of rHDL-aclacinomycin complexes that were intended for hepatoma chemotherapy [89]. The rHDL-aclacinomycin complexes were shown to have a molecular diameter and molecular weight similar to native HDL [89]. Lou *et al.* also

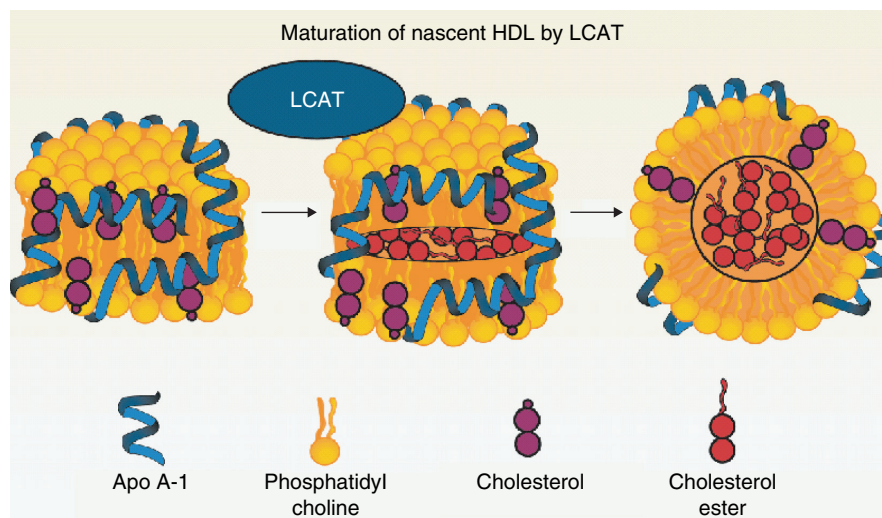


Figure 2. Transformation of the discoidal precursor (nascent) HDL to its stable, spherical configuration via the accumulation of cholesteryl esters, catalyzed by lecithin–cholesterol acyltransferase.

Reproduced from ALEXANDER ET, BHAT S, THOMAS MJ *et al.*: Apolipoprotein A-I helix 6 negatively charged residues attenuate lecithin–cholesterol acyltransferase (LCAT) reactivity. *Biochemistry* (2005) **44**(14):5409–5419.

Apo A-1: Apolipoprotein A-I; HDL: High-density lipoprotein; LCAT: Lecithin–cholesterol acyltransferase.

reported that the rHDL-aclacinomycin complex was taken up more avidly by hepatoma cells than normal cells, thus, providing preliminary evidence for the selective targeting potential of their formulation [89]. In another attempt to use synthetic HDL complexes for the delivery of pharmaceutical agents, Oda *et al.* incorporated amphotericin B into apolipoprotein A-I-containing discoidal particles that exhibited strong *in vitro* and *in vivo* antifungal activity [90].

Essentially, all the drugs that have been incorporated into LDL- or HDL-based vehicles, including those quoted in this review [12,15,88–90], are substantially lipophilic and, thus, resemble cholesteryl esters (the native lipoprotein core component; Figure 1) regarding their solubility properties. In the present authors' experience, a helpful way to define the potential to incorporate specific drugs into lipoprotein-type vehicles is to refer to their octanol/water partition coefficient (XlogP) [91]. The respective values for most compounds, including anti-cancer drugs may be found at [301]. In the present authors' opinion, compounds with an XlogP value > 1 are reasonable candidates for incorporation into synthetic HDL nanoparticles. The tendency to be incorporated into the core of the lipoprotein particles improves as the XlogP value exceeds 3.

3.3 The efficacy of high-density lipoprotein-type carriers for anti-cancer agents

The targeted delivery of anti-cancer agents via lipoprotein carriers is based on the concept that cancer cells and tumors have a higher expression of lipoprotein receptors [9,14,18,27], due to their increased need for cholesterol to promote rapid proliferation [64]. Clinical studies have shown that HDL cholesterol levels, as with LDL cholesterol, were lower

in a group of cancer patients than in normal controls [66–70]. Recent animal studies are in agreement with the above clinical data showing that the HDL receptors were over-expressed in breast cancer cells compared with surrounding normal cells [17]. In addition, Pussinen *et al.* [11] have shown earlier that breast cancer cells acquired cholesteryl esters from HDL via the CLA-1 receptor, indicating that HDL receptors may play an essential role in regulating the growth of cancer cells/tumors and, thus, support the hypothesis that the selective targeting of anti-cancer agents delivered via HDL-type formulations is feasible [27]. Earlier findings from the present authors' laboratory showed that cancer cells produced substantially stronger immunoblots with an scavenger receptor B1 antibody than normal fibroblasts, and that the respective uptakes of paclitaxel and cholesteryl esters were highly correlated when delivered by rHDL nanoparticles, suggesting a receptor-mediated pathway for the entry of paclitaxel into cancer cells [12].

4. Enhanced targeting of lipoprotein–drug complexes via covalent modification

Although, as emphasized earlier, the receptor-mediated uptake of drugs from lipoprotein complexes by cancer cells and tumors is a major advantage over other types of formulations, the interaction of lipoprotein nanoparticles with normal tissues resulting in side effects due to the enhanced delivery of a cytotoxic agent is a potential concern. A recent strategy developed by Zheng *et al.* [92] addressed this problem by masking the native receptor interaction between the lipoprotein and its target cells via attaching specific surface ligands to the lipoprotein–drug

complex with exceptionally high affinity for cancer cells and presumably malignant tissues. This strategy is based on the hypothesis that the delivery of anti-cancer drugs to cancer cells and tumors can be substantially enhanced by the interaction between a homing molecule (e.g., folate, antibodies) that is attached to the lipoprotein carrier. Such an approach could provide essentially unlimited opportunities for specifically targeting lipoprotein/drug complexes to individual types of malignant tumors and, thus, could revolutionize cancer chemotherapy.

5. Comparison of lipoprotein-mediated drug delivery with other formulations

Due to the tremendous variety of drug delivery formulations, it is only possible to evaluate a limited number of these. The major emphasis, therefore, will be on selected applications of liposomes, lipophilic pro-drugs and serum albumin-containing formulations using the application of paclitaxel in cancer chemotherapy as a model. Paclitaxel is a hydrophobic compound that requires emulsification/solubilization for intravenous administration [92]. The most widely used formulation (Taxol) contains the emulsifier Cremophor EL® (BASF), which has an undesirable impact in chemotherapy, due to numerous side effects [92,93]. Since the initial marketing of Taxol, several alternate formulations were developed, primarily to eliminate the Cremophor component and to increase the solubility and targeting of paclitaxel. Among the major preparations containing paclitaxel are liposomal formulations, nanoparticles and microemulsions [94-99], prodrugs, antibody conjugates [100-106] and an albumin complex [107,108] that for the most part have been shown to have superior biological activity and anti-tumor potential to Taxol.

These lipoprotein-based formulations offer potentially enhanced performance over the above alternatives, due to their natural components that render them biodegradable and help to avoid rapid removal by the reticuloendothelial system [24,50]. The biological compatibility and safety [24,48,50] are also important issues that favor lipoproteins over other delivery systems, as several formulations containing HDL components have been already been safely injected into human subjects [109-114]. Lipoprotein formulations tend to have substantially smaller molecular diameters than most liposomal drug preparations. This may be a considerable advantage, as smaller liposomes have been considered to be superior drug delivery agents based on their pharmacokinetic properties [115]. Another important issue favoring lipoproteins over other formulations is targeting, as both LDL and HDL drug complexes are known to deliver their pharmaceutical load via receptor-mediated mechanisms that are known to be overexpressed in cancer cells compared with normal cells [9,12,17,27]. This 'magic bullet'-like capability [18] applies particularly to the comparison of lipoproteins to one of the recently developed paclitaxel formulations. Abraxane or ABI-007, a paclitaxel serum albumin complex, has been

shown to be considerably more effective than Taxol due to an increase in the maximum tolerated dose and the reduction of side effects, particularly hypersensitivity [107,108]. Nevertheless, it appears that additional therapeutic benefits may accrue via the lipoprotein delivery of paclitaxel and other hydrophobic drugs, as Edwards *et al.* have shown that the delivery of omega-3 fatty acids to cancer cells produced markedly stronger inhibitory and apoptotic responses when facilitated by LDL compared with serum albumin [116].

6. Conclusions

Despite many potential advantages over other drug delivery systems, the therapeutic applications of lipoproteins and synthetic lipid-protein complexes resembling lipoproteins continues to lag behind all other formulations. Although lipoprotein preparations are biologically compatible and offer superb targeting potential via receptor-mediated mechanisms, the difficulty in obtaining the polypeptide starting material has so far been a major obstacle in the development of lipoprotein-based pharmaceutical formulations. Perhaps the use of peptide fragments, representing the receptor binding regions of specific apolipoproteins, may be useful. This approach would allow the design and preparation of synthetic lipoproteins to facilitate the scaling up of this process to the production level and allow the pharmaceutical manufacture of lipoprotein-drug formulations.

7. Expert opinion

Pharmaceutical formulations utilizing native and especially synthetic lipoproteins have the potential to perform favorably compared with most alternative drug delivery systems, especially for cancer chemotherapy. The main advantages in favor of lipoprotein-based formulations are their functional capacity to transport hydrophobic drugs, their biocompatible components, relative stability in the blood circulation and their track record of having already been safely injected into human subjects [109-114]. Regarding their utilization in cancer chemotherapy, an additional major advantage favoring lipoprotein-based formulations is their vast targeting potential, via receptor-mediated mechanisms that are overexpressed in cancer cells versus normal cells. Furthermore, an enhanced targeting strategy may be employed by attaching ligands to the apolipoprotein component(s) of the lipoprotein complex to home in on overexpressed surface antigens that characterize specific cancer cells and tumors. This type of approach could eventually lead to individualized therapy and, thus, revolutionize cancer treatment.

Although nearly 25 years have passed since Counsell and Pohland first proposed the use of lipoproteins as delivery vehicles for diagnostic and therapeutic agents [13], there are presently no lipoprotein-based formulations in clinical use or in clinical trials. Since the original proposal to utilize native or synthetic lipoproteins as drug delivery vehicles,

several patent applications have been filed to protect inventions in this area [117]. Some of these involve the use of peptide fragments, representing the receptor binding regions or other specific domains of apolipoproteins. Using this approach may allow the efficient production of synthetic lipoproteins and its scaling up to the industrial level.

From the financial/business point of view, an additional advantage in the utilization of lipoprotein type carriers as drug delivery agents lies in the potential for not only developing formulations with emerging pharmaceuticals, but the revitalization of numerous highly efficient compounds

that had been set aside due to their poor solubility properties. Nevertheless, although the scientific data predict a bright future, until a major commitment is forthcoming from pharmaceutical companies to facilitate their development, lipoprotein-based formulations may remain the best kept secret in the drug delivery arena.

Declaration of interest

The authors state no conflict of interest and have received no payment in preparation of this manuscript.

Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- ROSEN H, ABRIBAT T: The rise and rise of drug delivery. *Nat. Rev. Drug Discov.* (2005) 4(5):381-385.
- MINCHINTON AI, TANNOK IF: Drug penetration in solid tumours. *Nat. Rev. Cancer* (2006) 6(8):583-592.
- YIH TC, LI-FANDI M: Engineered nanoparticles as precise drug delivery systems. *J. Cell Biochem.* (2006) 97(6):1184-1190.
- COX MC, DAN TD, SWAIN SM: Emerging drugs to replace current leaders in first-line therapy for breast cancer. *Expert Opin. Emerging Drugs* (2006) 11(3):489-501.
- BLAY JY, LE CESNE A, ALBERTI L, RAY-COQUART I: Targeted cancer therapies. *Bull. Cancer* (2005) 92(2):E13-E18.
- TULENKO TN, SUMNER AE: The physiology of lipoproteins. *J. Nucl. Cardiol.* (2002) 9(6):638-649.
- HEVONOJA T, PENTIKAINEN MO, HYVONEN MT, KOVANEN PT, ALA-KORPELA M: Structure of low density lipoprotein (LDL) particles: basis for understanding molecular changes in modified LDL. *Biochim. Biophys. Acta* (2000) 1488(3):189-210.
- WANG M, BRIGGS MR: HDL: The metabolism, function, and therapeutic importance. *Chem. Rev.* (2004) 104:119-137.
- HO YK, SMITH RG, BROWN MS, GOLDSTEIN JL: Low-density lipoprotein (LDL) receptor activity in human acute myelogenous leukemia cells. *Blood* (1978) 52(6):1099-1114.
- KRIEGER M, SMITH LC, ANDERSON RG *et al.*: Reconstituted low density lipoprotein: a vehicle for the delivery of hydrophobic fluorescent probes to cells. *J. Supramol. Struct.* (1979) 10(4):467-478.
- PUSSINEN PJ, KARTEN B, WINTERSPERGER A *et al.*: The human breast carcinoma cell line HBL-100 acquires exogenous cholesterol from high-density lipoprotein via CLA-1 (CD-36 and LIMPII analogous 1)-mediated selective cholesteryl ester uptake. *Biochem. J.* (2000) 349(2):559-566.
- LACKO AG, NAIR M, PARANJAPPE S, JOHNSON S, MCCONATHY WJ: High density lipoprotein complexes as delivery vehicles for anticancer drugs. *Anticancer Res.* (2002) 22(4):2045-2049.
- COUNSELL RE, POHLAND RC: Lipoproteins as potential site-specific delivery systems for diagnostic and therapeutic agents. *J. Med. Chem.* (1982) 25(10):111-125.
- This is the first report to recommend lipoproteins as drug delivery agents.
- IMACHI H, MURAO K, SAYO Y *et al.*: Evidence for a potential role for HDL as an important source of cholesterol in human adrenocortical tumors via the CLA-1 pathway. *Endocr. J.* (1999) 46(1):27-34.
- GAL D, OHASHI M, MACDONALD PC, BUCHSBAUM HJ, SIMPSON ER: Low-density lipoprotein as a potential vehicle for chemotherapeutic agents and radionucleotides in the management of gynecologic neoplasms. *Am. J. Obstet. Gynecol.* (1981) 139(8):877-885.
- LUNDBERG B: Preparation of drug-low density lipoprotein complexes for delivery of antitumoral drugs via the low density lipoprotein pathway. *Cancer Res.* (1987) 47(15):4105-4108.
- CAO WM, MURAO K, IMACHI H *et al.*: A mutant high-density lipoprotein receptor inhibits proliferation of human breast cancer cells. *Cancer Res.* (2004) 64(4):1515-1521.
- This study clearly demonstrates the upregulation of HDL receptors in cancer cells.
- LACKO AG, NAIR M, PARANJAPPE S, MCCONATHY WJ: Trojan horse meets magic bullet to spawn a novel, highly effective drug delivery model. *Chemotherapy* (2006) 52(4):171-173.
- KRIEGER M, BROWN MS, FAUST JR, GOLDSTEIN JL: Replacement of endogenous cholesteryl esters of low density lipoprotein with exogenous cholesteryl linoleate. Reconstitution of a biologically active lipoprotein particle. *J. Biol. Chem.* (1978) 253(12):4093-4101.
- MASQUELIER M, TIRZITIS G, PETERSON CO *et al.*: Plasma stability and cytotoxicity of lipophilic daunorubicin derivatives incorporated into low density lipoproteins. *Eur. J. Med. Chem.* (2000) 35(40):429-438.
- MASQUELIER M, LUNDBERG B, PETERSON CO, VITOLS SG: Cytotoxic effect of a lipophilic alkylating agent after incorporation into low density lipoprotein or emulsions: studies in human leukemic cells. *Leuk. Res.* (2006) 30(2):136-144.
- TATIDIS L, MASQUELIER M, VITOLS SG: Elevated uptake of low density lipoprotein by drug resistant human leukemic cell lines. *Biochem. Pharmacol.* (2002) 63(12):2169-2180.
- KADER A, PATER AJ: Loading anticancer drugs into HDL as well as LDL has little affect on properties of complexes and enhances cytotoxicity to human carcinoma cells. *J. Control. Rel.* (2002) 80(1-3):29-44.

24. DE SMIDT PC, VAN BERKEL TJ: LDL-mediated drug targeting. *Crit. Rev. Ther. Drug Carrier Syst.* (1990) 7(2):99-120.
- **This is an excellent review evaluating the pros and cons of lipoproteins up to that date.**
25. VITOLS S, GAHRTON G, OST A, PETERSON C: Elevated low density lipoprotein receptor activity in leukemic cells with monocytic differentiation. *Blood* (1984) 63(5):1186-1193.
26. GAL D, MACDONALD PC, PORTER JC, SIMPSON ER: Cholesterol metabolism in cancer cells in monolayer culture. III. Low-density lipoprotein metabolism. *Int. J. Cancer* (1981) 28(3):315-319.
27. VITOLS S, ANGELIN B, ERICSSON S *et al.*: Uptake of low density lipoproteins by human leukemic cells *in vivo*: relation to plasma lipoprotein levels and possible relevance for selective chemotherapy. *Proc. Natl. Acad. Sci. USA* (1990) 87(7):2598-2602.
- **This paper presents a clear rationale for lipoproteins as potentially targeted delivery vehicles.**
28. CLAYMAN RV, BILHARTZ LE, SPADY DK, BUJA LM, DIETSCHY JM: Low density lipoprotein-receptor activity is lost *in vivo* in malignantly transformed renal tissue. *FEBS Lett.* (1986) 196(1):87-90.
29. FABRICANT M, BROITMAN SA: Evidence for deficiency of low density lipoprotein receptor on human colonic carcinoma cell lines. *Cancer Res.* (1990) 50(3):632-636.
30. ANDERSON RG, BROWN MS, GOLDSTEIN JL: Inefficient internalization of receptor-bound low density lipoprotein in human carcinoma A-431 cells. *J. Cell Biol.* (1981) 88(2):441-452.
31. RUDLING M, GAFVELS M, PARINI P, GAHRTON G, ANGELIN B: Lipoprotein receptors in acute myelogenous leukemia: failure to detect increased low-density lipoprotein (LDL) receptor numbers in cell membranes despite increased cellular LDL degradation. *Am. J. Pathol.* (1998) 153(2):1923-1935.
32. MASQUELIER M, VITOLS S, PETERSON C: Low-density lipoprotein as a carrier of antitumoral drugs: *in vivo* fate of drug-human low-density lipoprotein complexes in mice. *Cancer Res.* (1986) 46(8):3842-3847.
33. VITOLS S, GAHRTON G, PETERSON C: Significance of the low-density lipoprotein (LDL) receptor pathway for the *in vitro* accumulation of AD-32 incorporated into LDL in normal and leukemic white blood cells. *Cancer Treat. Rep.* (1984) 68(3):515-520.
34. VITOLS S, SÖDERBERG-REID K, MASQUELIER M, SJÖSTRÖM B, PETERSON C: Low density lipoprotein for delivery of a water-insoluble alkylating agent to malignant cells. *In vitro* and *in vivo* studies of a drug-lipoprotein complex. *Br. J. Cancer* (1990) 62(5):724-729.
35. TATIDIS L, MASQUELIER M, VITOLS S: Elevated uptake of low density lipoprotein by drug resistant human leukemic cell lines. *Biochem. Pharmacol.* (2002) 63(12):2169-2180.
36. MASQUELIER M, VITOLS S, PALSSON M, MARS U, LARSSON BS, PETERSON C: Low density lipoprotein as a carrier of cytostatics in cancer chemotherapy: study of stability of drug-carrier complexes in blood. *J. Drug Target.* (2000) 8(3):155-164.
37. VITOLS S, MASQUELIER M, PETERSON C: Selective uptake of a toxic lipophilic anthracycline derivative by the low-density lipoprotein receptor pathway in cultured fibroblasts. *J. Med. Chem.* (1985) 28(4):451-454.
38. MASQUELIER M, TIRIYIS G, PETERSON C *et al.*: Plasma stability and cytotoxicity of lipophilic daunorubicin derivatives incorporated into low density lipoproteins. *Eur. J. Med. Chem.* (2000) 35(4):429-438.
39. LO EH, OOI VE, FUNG KP: Circumvention of multidrug resistance and reduction of cardiotoxicity of doxorubicin *in vivo* by coupling it with low density lipoprotein. *Life Sci.* (2002) 72(6):677-687.
40. CHU AC, TSANG SY, LO EH, FUNG KP: Plasma stability and cytotoxicity of lipophilic daunorubicin derivatives incorporated into low density lipoproteins. *Life Sci.* (2001) 70(5):591-601.
41. JORI G, REDDI E: The role of lipoproteins in the delivery of tumour-targeting photosensitizers. *Int. J. Biochem.* (1993) 25(10):1369-1375.
42. ALLISON BA, PRITCHARD PH, LEVY JG: Evidence for low-density lipoprotein receptor-mediated uptake of benzoporphyrin derivative. *Br. J. Cancer* (1994) 69(5):833-839.
43. HAMMEL M, LAGGNER P, PRASSL R: Structural characterisation of nucleoside loaded low density lipoprotein as a main criterion for the applicability as drug delivery system. *Chem. Phys. Lipids* (2003) 123(2):193-207.
44. MENG QH, WAHALA K, ADLERCREUTZ H, TIKKANEN MJ: Antiproliferative efficacy of lipophilic soy isoflavone phytoestrogens delivered by low density lipoprotein particles into cultured U937 cells. *Life Sci.* (1999) 65(16):1695-1705.
45. LESTAVEL-DELATTRE S, MARTIN-NIZARD F, CLAVEY V *et al.*: Low-density lipoprotein for delivery of an acrylophenone antineoplastic molecule into malignant cells. *Cancer Res.* (1992) 52(13):3629-3635.
46. SQALLI-HOUSSAIN H, PIERLOT C, KUSNIERZ JP *et al.*: Preparation of anti-HIV-low-density lipoprotein complexes for delivery of anti-HIV drugs via the low-density lipoprotein pathways. *Biotechnol. Ther.* (1994) 5:69-85.
47. LI H, GRAY BD, CORBIN I *et al.*: MR and fluorescent imaging of low-density lipoprotein receptors. *Acad. Radiol.* (2004) 11(11):1251-1259.
48. ADAMS T, ALANAZI F, LU DR: Safety and utilization of blood components as therapeutic delivery systems. *Curr. Pharm. Biotechnol.* (2003) 4(5):275-282.
49. OWENS MD, BAILLIE G, HALBERT GW: Physicochemical properties of microemulsion analogues of low density lipoprotein containing amphiphatic apolipoprotein B receptor sequences. *Int. J. Pharm.* (2001) 228(1-2):109-117.
- **This is the first attempt to use artificial surrogates for apolipoproteins as essential components for lipoprotein-based drug delivery vehicles.**
50. SHAW JM, SHAW KV: Key issues in the delivery of pharmacological agents using lipoproteins: design of a synthetic apolipoprotein-lipid carrier. *Target. Diagn. Ther.* (1991) 5:351-383.
51. WALSH MT, ATKINSON D: Reassembly of low-density lipoproteins. *Methods Enzymol.* (1986) 128:582-608.

52. RENSEN PC, DE VRUEH RL, KUIPER J, BIJSTERBOSCH MK, BIESSEN EA, VAN BERKEL TJ: Recombinant lipoproteins: lipoprotein-like lipid particles for drug targeting. *Adv. Drug Deliv. Rev.* (2001) 47(2-3):251-276.
53. LUNDBERG B, SUOMINEN L: Preparation of biologically active analogs of serum low density lipoprotein. *J. Lipid Res.* (1984) 25(6):550-558.
54. LUNDBERG B: Abstract cytotoxic activity of two new lipophilic steroid nitrogen carbamates incorporated into low-density lipoprotein. *Anticancer Drug Des.* (1994) 9(5):471-476.
55. RENSEN PC, SCHIFFELERS RM, VERSLUIS AJ, BIJSTERBOSCH MK, VAN KUIJK-MEUWISSEN ME, VAN BERKEL TJ: Human recombinant apolipoprotein E-enriched liposomes can mimic low-density lipoproteins as carriers for the site-specific delivery of antitumor agents. *Mol. Pharmacol.* (1997) 52(3):445-455.
56. VERSLUIS AJ, RENSEN PC, RUMP ET, VAN BERKEL TJ, BIJSTERBOSCH MK: Low-density lipoprotein receptor-mediated delivery of a lipophilic daunorubicin derivative to B16 tumours in mice using apolipoprotein E-enriched liposomes. *Br. J. Cancer* (1998) 78(12):1607-1614.
57. MARANHÃO RC, CESAR TB, PEDROSO-MARIANI SR, HIRATA MH, MESQUITA CH: Metabolic behavior in rats of a nonprotein microemulsion resembling low-density lipoprotein. *Lipids* (1993) 28(8):691-696.
58. LO PRETE AC, MARIA DA, RODRIGUES DG, VALDUGA CJ, IBANEZ OC, MARANHÃO RC: Evaluation in melanoma-bearing mice of an etoposide derivative associated to a cholesterol-rich nano-emulsion. *J. Pharm. Pharmacol.* (2006) 58(6):801-808.
59. RODRIGUES DG, MARIA DA, FERNANDES DC *et al.*: Improvement of paclitaxel therapeutic index by derivatization and association to a cholesterol-rich microemulsion: *in vitro* and *in vivo* studies. *Cancer Chemother. Pharmacol.* (2005) 55(6):565-576.
60. DIAS ML, CARVALHO JP, RODRIGUES DG, GRAZIANI SR, MARANHÃO RC: Pharmacokinetics and tumor uptake of a derivatized form of paclitaxel associated to a cholesterol-rich nanoemulsion (LDE) in patients with gynecologic cancers. *Cancer Chemother. Pharmacol.* (2007) 59(1):105-111.
61. BAILLIE G, OWENS MD, HALBERT GW: A synthetic low density lipoprotein particle capable of supporting U937 proliferation *in vitro*. *J. Lipid Res.* (2002) 43(1):69-73.
62. NIKANJAM M, BLAKELY EA, BJORNSTAD KA, SHU X, BUDINGER TF, FORTE TM: Synthetic nano-low density lipoprotein as targeted drug delivery vehicle for glioblastoma multiforme. *Int. J. Pharm.* (2006) 328(1):86-94.
63. BROWN MS, GOLDSTEIN JL: A receptor-mediated pathway for cholesterol homeostasis. *Science* (1986) 232(4746):34-47.
64. MARKEL A, BROOK GJ: Cancer and hypocholesterolemia. *Isr. J. Med. Sci.* (1994) 30(10):787-793.
65. FIORENZA AM, BRANCHI A, SOMMARIVA D: Serum lipoprotein profile in patients with cancer. A comparison with non-cancer subjects. *Int. J. Clin. Lab. Res.* (2000) 30(3):141-145.
66. DESSI S, BATETTA B, PULISCI D, ACCOGLI P, PANI P, BROCCIA G: Total and HDL cholesterol in human hematologic neoplasms. *Int. J. Hematol.* (1991) 54(6):483-486.
67. MOSCHOVI M, TRIMIS G, APOSTOLAKOU F, PAPASSOTIRIOU I, TZORZATOU-STATHOPOLOU F: Serum lipid alterations in acute lymphoblastic leukemia of childhood. *Pediatr. Hematol. Oncol.* (2004) 26(5):289-293.
68. SCRIBANO D, BARONI S, PAGANO L, ZUPPI C, LEONE G, GIARDINA B: Return to normal values of lipid pattern after effective chemotherapy in acute lymphoblastic leukemia. *Haematologica* (1996) 81(4):343-345.
69. SIEMIANOVICZ K, GMINSKI J, STAJSZCZYK M *et al.*: Serum LDL cholesterol concentration and lipoprotein electrophoresis pattern in patients with small cell lung cancer. *Int. J. Mol. Med.* (2000) 6(3):307-311.
70. BAYERDÖRFER E, MANNES GA, RICHTER WO *et al.*: Decreased high-density lipoprotein cholesterol and increased low-density cholesterol levels in patients with colorectal adenomas. *Ann. Intern. Med.* (1993) 118(7):481-487.
71. FIDGE NH: High density lipoprotein receptors, binding proteins, and ligands. *J. Lipid Res.* (1999) 40(2):187-201.
72. MURAO K, TERPSTRA V, GREEN SR, KONDRATENKO N, STEINBERG D, QUEHENBERGER O: Characterization of CLA-1, a human homologue of rodent scavenger receptor BI, as a receptor for high density lipoprotein and apoptotic thymocytes. *J. Biol. Chem.* (1997) 272(28):17551-17557.
73. RIGOTTI A, KRIEGER M: Getting a handle on "good" cholesterol with the high-density lipoprotein receptor. *N. Engl. J. Med.* (1999) 341(26):2011-2013.
74. FAVRE G, TAZI KA, LE GAILLARD F, BENNIS F, HACHEM H, SOULA G: High density lipoprotein-3 binding sites are related to DNA biosynthesis in the adenocarcinoma cell line A549. *J. Lipid Res.* (1993) 34(7):1093-1106.
75. GOSPODAROWICZ D, LUI GM, GONZALEZ R: High-density lipoproteins and the proliferation of human tumor cells maintained on extracellular matrix-coated dishes and exposed to defined medium. *Cancer Res.* (1982) 42(9):3704-3713.
76. JOZAN S, FAYE JC, TOURNIER JF, TAUBER JP, DAVID JF, BAYARD F: Interaction of estradiol and high density lipoproteins on proliferation of the human breast cancer cell line MCF-7 adapted to grow in serum free conditions. *Biochem. Biophys. Res. Commun.* (1985) 133(1):105-112.
77. WADSACK C, HIRSCHMUGL B, HAMMER A *et al.*: Scavenger receptor class B, type I on non-malignant and malignant human epithelial cells mediates cholesteryl ester-uptake from high density lipoproteins. *Int. J. Biochem. Cell Biol.* (2003) 35(4):441-454.
78. WADSACK C, HRZENJAK A, HAMMER A *et al.*: Trophoblast-like human choriocarcinoma cells serve as a suitable *in vitro* model for selective cholesteryl ester uptake from high density lipoproteins. *Eur. J. Biochem.* (2003) 270(3):451-462.
79. BARTER PJ: Hugh sinclair lecture: the regulation and remodelling of HDL by

- plasma factors. *Atheroscler. Suppl.* (2002) 3(4):39-47.
80. LINSEL-NITSCHKE P, TALL AR: HDL as a target in the treatment of atherosclerotic cardiovascular disease. *Nat. Rev. Drug Discov.* (2005) 4(3):193-205.
 81. HAMILTON RL, WILLIAMS MC, FIELDING CJ, HAVEL RJ: Discoidal bilayer structure of nascent high density lipoproteins from perfused rat liver. *J. Clin. Invest.* (1976) 58(3):667-680.
 82. FORTE T, NORUM KR, GLOMSET JA, NICHOLS AV: Plasma lipoproteins in familial lecithin: cholesterol acyltransferase deficiency: structure of low and high density lipoproteins as revealed by electron microscopy. *J. Clin. Invest.* (1971) 50(5):1141-1148.
 83. LERCH PG, FORTSCH V, HODLER G, BOLLI R: Production and characterization of a reconstituted high density lipoprotein for therapeutic applications. *Vox. Sang.* (1996) 71(3):155-164.
 84. PARKER TS, LEVINE DM, CHANG JC, LAXER J, COFFIN CC, RUBIN AL: Reconstituted high-density lipoprotein neutralizes gram-negative bacterial lipopolysaccharides in human whole blood. *Infect. Immun.* (1995) 63(1):253-258.
 85. SPARKS DL, PHILLIPS MC, LUND-KATZ SJ: The conformation of apolipoprotein A-I in discoidal and spherical recombinant high density lipoprotein particles. ^{13}C NMR studies of lysine ionization behavior. *J. Biol. Chem.* (1992) 267(36):25839-25847.
 86. JONAS A, WALD JH, TOOHILL KL, KRUL ES, KEZDY KE: Apolipoprotein A-I structure and lipid properties in homogeneous, reconstituted spherical and discoidal high density lipoproteins. *J. Biol. Chem.* (1990) 265(36):22123-22129.
 87. SCHOUTEN D, VAN DER KOOIJ M, MULLER J, PIETERS MN, BIJSTERBOSCH MK, VAN BERKEL TJ: Development of lipoprotein-like lipid particles for drug targeting: neo-high density lipoproteins. *Mol. Pharmacol.* (1993) 44(2):486-492.
 - **This is a good review summarizing the properties and the capabilities of synthetic HDL drug delivery vehicles.**
 88. BIJSTERBOSCH MK, SCHOUTEN D, VAN BERKEL TJ: Synthesis of the dioleoyl derivative of iododeoxyuridine and its incorporation into reconstituted high density lipoprotein particles. *Biochemistry* (1994) 33(47):14073-14080.
 89. LOU B, LIAO XL, WU MP, CHENG PF, YIN CY, FEI Z: High-density lipoprotein as a potential carrier for delivery of a lipophilic antitumoral drug into hepatoma cells. *World J. Gastroenterol.* (2005) 11(7):954-959.
 90. ODA MN, HARGREAVES PL, BECKSTEAD JA, REDMOND KA, VAN ANTWERPEN R, RYAN RO: Reconstituted high density lipoprotein enriched with the polyene antibiotic amphotericin B. *J. Lipid Res.* (2006) 47(2):260-267.
 91. WANG R, FU Y, LAI L: A new atom-additive method for calculating partition coefficients. *J. Chem. Inf. Comput. Sci.* (1997) 37:615-621.
 92. ZHENG G, CHEN J, LI H, GLICKSON JD: Rerouting lipoprotein nanoparticles to selected alternate receptors for the targeted delivery of cancer diagnostic and therapeutic agents. *Proc. Natl. Acad. Sci. USA* (2005) 102(49):17757-17762.
 - **This is a landmark paper demonstrating the effectiveness of enhanced targeting via the chemical modification of lipoprotein carriers.**
 93. NUJEN B, BOUMA M, SCHELLEN JH, BEIJNEN JH: Progress in the development of alternative pharmaceutical formulations of taxanes. *Invest. New Drugs* (2001) 19(2):143-53.
 93. SINGLA AK, GARG A, AGGARWAL D: Paclitaxel and its formulations. *Int. J. Pharm.* (2002) 235(1-2):179-192.
 94. TORCHILIN VP: Fluorescence microscopy to follow the targeting of liposomes and micelles to cells and their intracellular fate. *Adv. Drug Deliv. Rev.* (2005) 57(1):95-109.
 95. STRIETH S, EICHHORN ME, SAUER B *et al.*: Neovascular targeting chemotherapy: encapsulation of paclitaxel in cationic liposomes impairs functional tumor microvasculature. *Int. J. Cancer* (2004) 110(1):117-124.
 96. FETTERLY GJ, STRAUBINGER RM: Pharmacokinetics of paclitaxel-containing liposomes in rats. *AAPS Pharm. Sci.* (2003) 5(4):E32.
 97. MU L, FENG SS: A novel controlled release formulation for the anticancer drug paclitaxel (Taxol): PLGA nanoparticles containing vitamin E TPGS. *J. Control. Rel.* (2003) 86(1):33-48.
 98. GUILLEMARD V, SARAGOVIC HU: Taxane-antibody conjugates afford potent cytotoxicity, enhanced solubility, and tumor target selectivity. *Cancer Res.* (2001) 61(2):694-699.
 99. LUNDBERG BB: A submicron lipid emulsion coated with amphipathic polyethylene glycol for parenteral administration of paclitaxel (Taxol). *J. Pharm. Pharmacol.* (1997) 49(1):16-21.
 100. CHORDIA MD, YUAN H, JAGTAP PG *et al.*: Synthesis and bioactivity of 2,4-diacetyl analogues of paclitaxel. *Bioorg. Med. Chem.* (2001) 9(1):171-178.
 101. DE GROOT FM, VAN BERKOM LW, SCHEEREN HW: Synthesis and biological evaluation of 2'-carbamate-linked and 2'-carbonate-linked prodrugs of paclitaxel: selective activation by the tumor-associated protease plasmin. *J. Med. Chem.* (2000) 43(16):3093-3102.
 102. SAFAVY A, BONNER JA, WAKSAL HW *et al.*: Synthesis and biological evaluation of paclitaxel-C225 conjugate as a model for targeted drug delivery. *Bioconjug. Chem.* (2003) 14(2):302-310.
 103. LEE JW, LU JY, LOW PS, FUCHS PL: Synthesis and evaluation of Taxol-folic acid conjugates as targeted antineoplastics. *Bioorg. Med. Chem.* (2002) 10(7):2397-2414.
 104. BRADLEY MO, WEBB NL, ANTHONY FH *et al.*: Tumor targeting by covalent conjugation of a natural fatty acid to paclitaxel. *Clin. Cancer Res.* (2001) 7(10):3229-3238.
 105. SUGAHARA S, KAJIKIM, KURIYAMA H, KOBAYASHI TR: Paclitaxel delivery systems: the use of amino acid linkers in the conjugation of paclitaxel with carboxymethyl dextran to create prodrugs. *Biol. Pharm. Bull.* (2002) 25(5):632-641.
 106. WU X, OJIMA I: Tumor specific novel taxoid-monoclonal antibody conjugates. *Curr. Med. Chem.* (2004) 11(4):429-438.
 107. IBRAHIM NK, DESAI N, LEGHA S *et al.*: Phase I and pharmacokinetic study of ABI-007, a cremophor-free, protein-stabilized, nanoparticle formulation of paclitaxel. *Clin. Cancer Res.* (2002) 8(5):1038-1044.
 108. DAMASCELLI B, CANTU G, MATTAVELLI F *et al.*: Intraarterial chemotherapy with polyoxyethylated

- castor oil free paclitaxel, incorporated in albumin nanoparticles (ABI-007): Phase II study of patients with squamous cell carcinoma of the head and neck and anal canal: preliminary evidence of clinical activity. *Cancer* (2001) 92(10):2592-2602.
109. BISOENDIAL RJ, HOVINGH GK, LEVELS JH *et al.*: Restoration of endothelial function by increasing high-density lipoprotein in subjects with isolated low high-density lipoprotein. *Circulation* (2003) 107(23):2944-2948.
 110. NANJEE MN, CROUSE JR, KING JM *et al.*: Effects of intravenous infusion of lipid-free apo A-I in humans. *Arterioscler. Thromb. Vasc. Biol.* (1996) 16(9):1203-1214.
 111. SPIEKER LE, SUDANO I, HÜRLIMANN D *et al.*: High-density lipoprotein restores endothelial function in hypercholesterolemic men. *Circulation* (2002) 105(12):1399-1402.
 112. HOVORKA R, NANJEE MN, COOKE CJ, MILLER IP, OLSZEWSKI WL, MILLER NE: Mass kinetics of apolipoprotein A-I in interstitial fluid after administration of intravenous apolipoprotein A-I/lecithin discs in humans. *J. Lipid Res.* (2006) 47(5):975-981.
 113. NANJEE MN, CROUSE JR, KING JM *et al.*: Effects of intravenous infusion of lipid-free apo A-I in humans. *Arterioscler. Thromb. Vasc. Biol.* (1996) 16(9):1203-1214.
 114. PAJKRT D, DORAN JE, KOSTER F *et al.*: Antiinflammatory effects of reconstituted high-density lipoprotein during human endotoxemia. *J. Exp. Med.* (1996) 184(5):1601-1608.
 115. PALATINI P: Disposition kinetics of phospholipid liposomes. *Adv. Exp. Med. Biol.* (1992) 318:375-391.
 116. EDWARDS IJ, BERQUIN IM, SUN H *et al.*: Differential effects of delivery of ω -3 fatty acids to human cancer cells by low-density lipoproteins versus albumin. *Clin. Cancer Res.* (2004) 10(24):8275-8283.
 117. LACKO AG, STEWART DR, MCCLAIN R, PROKAI L, MCCONATHY WJ: Recent developments and patenting of lipoprotein based formulations. *Recent Patents on Drug Delivery Formulation* (2007) 1:143-145.

Patent

201. LEVINE DM, SIMON SR, GORDON BR, PARKER TS, SAAL SD, RUBIN AL: Reconstituted HDL particles and uses thereof. US5128318 (1992).

Website

301. <http://pubchem.ncbi.nlm.nih.gov> PubChem. Accessed 21st September 2007.

Affiliation

Andras G Lacko[†] PhD, Maya Nair PhD, Laszlo Prokai PhD & Walter J McConathy PhD
[†]Author for correspondence
 University of North Texas Health Science Center, Department of Molecular Biology and Immunology,
 3500 Camp Bowie Blvd,
 Fort Worth, TX 76107, USA
 Tel: +1 817 735 2132; Fax: +1 817 735 2118;
 E-mail: alacko@hsc.unt.edu

